

Experimental evidence for an alkali ecotype of *Lolium multiflorum*, an exotic invasive annual grass in the Central Valley, CA, USA

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Abstract One of the characteristics of highly invaded ecosystems is that exotic species are often poor invaders of edaphically severe sites, which become refuges for native flora. To investigate the invasive potential of *Lolium multiflorum* (Per.) into alkali sites in California, an ex-situ reciprocal transfer experiment was carried out using seeds from populations of *L. multiflorum* taken from three sites differing in alkalinity (and inundation), including alkali sink soils (pH 8.5) and sink matrix soils (pH 7.4) located within meters of each other, and non-sink soils (pH 5.0) located several km away. Survivorship, plant height, leaf number and seed production were assessed. In addition, a native composite, *Hemizonia pungens* (Hick.), commonly found on alkali sinks was also sampled at the sink and sink matrix microsites. *Lolium multiflorum* plants grown from alkali sink and sink

matrix seeds produced fewer leaves and seeds but were taller than plants grown from non-alkali seeds, the latter perhaps an adaptation to frequently inundated soils. Non-alkali genotypes fared poorly in sink soils for all traits, both in comparison to their growth on non-sink soils, and in comparison to the sink and sink edge genotypes. This suggests the existence of *L. multiflorum* ecotypes adapted to inundated alkali sinks, a genotypic difference that occurs on a broad spatial scale (kilometers), but not so obviously on the microsite scale (meters) between sink and sink matrix populations. These data suggest that the absence of exotic invasives from alkali sites may be temporary if they are evolving tolerance for these severe sites, and this may threaten the future of the native alkali specialists that currently find refuge in these sites.

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Introduction

Introduced plants have had devastating effects on local biodiversity worldwide (Vitousek et al. 1997; Wilcove et al. 1998). One way these alien species pose a serious threat is when they are competitively superior to indigenous species. When the local species and alien species have

evolved in allopatry, on exposure the indigenous species may lack the ability to successfully compete for light, space, or nutrients. The invasive on the other hand may find itself in a community that lacks similar competitors and therefore has the potential to successfully invade whole habitats.

In particular, the non-forested habitats of the western United States have undergone massive invasions by exotic invasive plant species. The grasslands, scrublands, and woodlands of interior California have had their herbaceous layers nearly completely replaced by invasive annuals (Heady 1988). These invasive exotic annual grasses are a limiting impediment to grassland restoration throughout the western United States (Kay et al. 1981; Dyer and Rice 1997, 1999; Brown and Rice 2000). However, within this matrix of exotic species are isolated communities that have resisted invasion. Edaphically extreme plant communities are characterized by a paucity of invaders, and serve as a refuge for native species. These edaphic refuges include serpentine outcrops (Harrison 1999; Gelbard and Harrison 2003; Gram et al. 2004; Brady et al. 2005) and vernal pools (Zedler and Black 2004; Marty 2005).

The alkali sinks of the Central Valley grasslands appear to be similar edaphic refuges. In some areas of impeded drainage, low-lying spots are prone to long-term inundation and are strongly alkaline. The plant communities of the background matrix are severely invaded by species such as *Lolium multiflorum* (Italian Ryegrass), which forms nearly monospecific stands. The sinks themselves are largely free of exotic plants and have considerable open ground and healthy stands of native alkali specialists. However, a few individuals of *L. multiflorum* are found in these sinks, and if they represent a vanguard of alkali ecotypes, may pose a long-term threat to native alkali biodiversity.

Research on native species has examined the role of genetic differentiation in ecotypic adaptation along stress gradients (e.g., Kruckeberg 1951; Bradshaw 1984; Bennington and McGraw 1995; Brady et al. 2005), and the particular life history, morphological and physiological traits responsible for fitness differences along stress gradients (Bennington and McGraw 1995; Brady et al. 2005). Although there has been recent

research documenting the (slow) invasion of edaphically severe sites by exotic species in both serpentine (Williamson and Harrison 2002) and vernal pool (Gerhardt and Collinge 2003; Zedler and Black 2004) refuges, there has been no research on the potential evolution of exotic ecotypes to the edaphically severe sites that have historically resisted their invasion.

Many introduced species respond genotypically to local environmental conditions and can evolve rapidly to produce an ecotype genetically distinct from their ancestors (Cox 2004). Annuals in particular may have an advantage in rapid specialization due to their short life cycles. Annual invasives with the potential to produce ecotypes rapidly may be a threat in edaphically extreme environments that support rare or endemic natives.

A standard method for investigating ecotypic traits of species is the reciprocal transplant technique (Leiss and Müller-Schärer 2001). If individuals grown in local conditions outperform individuals grown in non-local conditions (the “local vs. foreign” criterion of Kawecki and Ebert 2004), this is evidence for evolutionary adaptation to local conditions (Turesson 1922 [from Briggs and Walters 1984], Clausen et al. 1940; Cheplick 1988; Primack and Kang 1989; McGraw and Chapin 1989; Joshi et al. 2001; Galloway and Fenster 2000; Montalvo and Ellstrand 2000; Berglund et al. 2004).

The aim of this study was to investigate the possibility that *L. multiflorum* at alkali sink sites has locally adapted ecotypes and therefore has increased potential for invasion in these sites. We analyzed survivorship, growth and fecundity of the exotic invasive *Lolium multiflorum* and the native alkali specialist *Hemizonia pungens* ssp. *pungens* taken from alkali sink and sink matrix sites, and in the case of *L. multiflorum* also a non-alkaline site, on different source soils using a greenhouse garden reciprocal transfer experiment.

Study species, study sites, and methods

Study species

Lolium multiflorum is a non-native annual grass first introduced to the United States of America

by Spanish colonists in the late 1600s (Hannaway et al. 1999). Because it is considered a valuable pasture grass, numerous introductions have occurred over the past 300 years, and it has become well established across the continent. It is now considered a pest species in some localities (Hannaway et al. 1999), although it is still actively marketed, and is recommended by the USDA for use in rangelands. The California Exotic Pest Plant Council lists it as a potential threat to vernal pools, but lacks sufficient information for a more precise determination (California Exotic Pest Plant Council 1999). The pest status of *L. multiflorum* is compounded by its ability to evolve increased resistance to herbicides (Agriculture and Agri-food Canada 1999). Although *L. multiflorum* prefers well-drained fertile soil, it can survive in poorly drained soil and a period of flooding of up to 20 days if temperatures are low (Hannaway et al. 1999). It is tolerant of acidic to mildly alkaline soils (reported range of pH 5.0 to 7.8), and prefers non-alkaline soils (pH 5.5 to 7.5; Hannaway et al. 1999). *Lolium multiflorum* germinates in the late winter or early spring and thrives in cool wet conditions. Under ex-situ conditions *L. multiflorum* germinates readily without any specific pre-treatments (Hellmers and Ashby 1958; K. Dawson, unpublished data).

Hemizonia pungens (tar plant) is a composite, native to interior grasslands of the Pacific coast. The subspecies *H. pungens* ssp. *pungens* is an alkali endemic to the Central Valley of California (Hickman 1993). The seeds of *H. pungens* require cold stratification for germination (Dawson, unpublished data). We included it here as a species with a long evolutionary history in these alkali sinks, in contrast to *L. multiflorum*.

Study sites

Seeds of the two plant species and soils were collected from two sites near Davis, CA. The first is a seasonally inundated pasture located near Willow Slough in the central valley of Yolo County, CA approximately 10 km northwest of Davis (38°37'N, 121°43'W). The site is characterized by alkali pools that retain water well into the spring annual growing season. In 2005, it became the Alkali Grasslands Preserve. This is one

of the few remaining areas with alkali sinks in the region, and was selected for its proximity to the university. We sampled both the sink microsites and the non-sink (“sink matrix”) microsites. *Lolium multiflorum* and *Hemizonia pungens* occurred in both microsites, although *L. multiflorum* was noticeably sparser in the sinks and *H. pungens* noticeably sparser in the sink matrix. *Lolium multiflorum* seeds were also collected from a second site, Bobcat Ranch conservation area, a rangeland located on sandy, well-drained non-alkali soils approximately 30 km west of the alkali site (38°33'N, 122°03'W). This site was selected because it was a conservation site that was not highly disturbed, and also has large stands of *L. multiflorum*. Samples of the three soil types (two alkali microsites and one non-alkali site) were sent to DANR labs at University of California Davis, for chemical and textural analysis.

Reciprocal transfer

A common garden experiment was carried out under greenhouse conditions. Sufficient soil was gathered from the same field sites as the seeds of both species. The top 30 cm of soil with the upper layer of vegetation removed was collected from the alkali site in March 2004. The soil was taken from both alkali sinks and sink matrix microsites, based on pH and visibly different vegetation cover and composition. The pH was initially tested in the field to assist in field differentiation of these two microsites. Soil from the non-alkali Bobcat Ranch site was collected within the same month and again the top 30 cm was sampled. The soil was air-dried for 12 days. It was broken into smaller particles and sifted, first to 3 mm and then to 1 mm. The soil was then heat-treated at 60°C for 72 h to reduce chances of foreign seeds germinating in the soil during the study. The clay-like nature of the soil from the alkali sites and the small size of the planting pots impeded free drainage of the soil. Therefore, to retain the particle size produced by the grinding and sieving process and to emulate “natural” drainage conditions, a polymer solution of 0.02% in de-ionized water was added to the soil in the pots. The soil was then dried for a further 48 h at 60°C. The resulting soil still had slow drainage, but did not

pool deeply. The heat treatments probably killed or reduced soil biota, including mycorrhizae, but probably had little effect on soil chemistry.

Lolium multiflorum was grown in $4 \times 4 \times 7$ cm plastic tubes, and *H. pungens* was grown in tubes of $3 \times 3 \times 5$ cm. A randomized block arrangement was used to accommodate 10 replicates of each of the nine soil/seed combinations (three soil types \times three seed types) for *L. multiflorum* and the 4 soil/seed combinations (two soil types \times two seed types) for *H. pungens*. The *H. pungens* seed was sown on 10 March 2004 and the *L. multiflorum* on 1 April 2004. Seeds of both species had been collected one year prior and stored at room temperature. The *H. pungens* seeds were cold stratified for several days. All seeds used were soaked in a 10% bleach solution as a fungicide and then rinsed. To assure at least one seedling per pot, three seeds were sown in each *L. multiflorum* pot and five in each *H. pungens* pot. Seedling trays were placed under a mist bench in the greenhouse. Impeded drainage occurred in some (alkali) seedling pots initially but soil drained more effectively after two weeks. All trays were watered to keep the soil moist. Thinning as necessary was carried out on 20 April for the *H. pungens* and on 5 May for *L. multiflorum* to one seedling per planting tube. Five individuals of *L. multiflorum* (out of 90) and 11 individuals of *H. pungens* (out of 40) died before reaching maturity. For neither species was there a significant pattern in this mortality with respect to either seed source or soil source.

The height and number of leaves of each seedling were measured weekly. Height was recorded as the length from the base to the tip of the tallest tiller for the *L. multiflorum* and from the base of the stem to the apical bud for the *H. pungens*. At the end of the growth study (9 June) the *L. multiflorum* and *H. pungens* were moved from the mist bench to a non-misted table in the same greenhouse and only watered 3 times a week, to induce seed set. The number of seeds produced was counted for each individual of *L. multiflorum*. None of the *H. pungens* flowered before the end of the experiment. For analysis we used height and number of leaves produced during the vegetative stage and, for *L. multiflorum*,

number of seeds produced during the reproductive stage.

Statistical analyses

We tested the effects of seed source and soil source on plant height, number of leaves, and number of seeds produced (seed set). Seed set was log-transformed for normality. We first tested for local ecotypes by comparing sink and sink matrix seed sources on sink and sink matrix soils for both *H. pungens* and *L. multiflorum*, looking for both main effects and interactions. We then tested for ecotypic differentiation in *L. multiflorum* between the alkali sink site and the non-alkali site. Because there were significant effects of soil source, but not of seed source (see Table 2), we lumped the sink and sink matrix seed sources (“alkali”) when comparing to non-alkali seed sources, but included all three soil sources, again with interaction terms. We examined the relationships between seed set and plant height and leaf number using correlation analysis.

Results

Compared to the non-alkali soils, the sink and sink matrix soils were characterized by high pH, electro-conductivity, N, P, K, Na, Cl, HCO_3 , Boron, and clay content (Table 1). For most of these traits, the sink matrix soils were intermediate, with values much closer to those in the sink soils than to those in the non-alkali soils. The alkali soil values of pH, Na, Cl, and B are excessive by agricultural standards (Prasad and Power 1997).

Data for height and leaf number provide no evidence of a differentiation of sink and sink matrix ecotypes in *H. pungens* (Table 2). (*Hemizonia pungens* plants had not yet reproduced by the end of the experiment.) Individuals from both seed sources did grow 50% taller on sink matrix soil than on sink soil (9.1 vs. 6.2 cm, $P = 0.01$).

For *Lolium multiflorum*, analysis of only the sink and sink matrix data reveals an effect of soil source: individuals grown on sink matrix soils produced 60% more leaves than those on sink

Table 1 Results of analysis of soils collected from inside alkali sinks, from the matrix surrounding the sinks, and from the non-alkali site. Water depths were measured during the height of the rainy season in February 2006

Soil trait	Sink	Sink matrix	Non-alkali
pH	8.5	7.4	5.0
EC (dS/m)	2.87	2.11	0.21
Na (meq/L)	26.3	18.6	0.6
Cl (meq/L)	8.5	11.4	1.0
HCO ₃ (meq/L)	5.1	4.5	0.9
CO ₃ (meq/L)	1.4	< 0.1	< 0.1
SAR	41	35	1
B (mg/L)	39.1	19.9	0.1
NO ₃ -N (ppm)	4.3	3.1	0.2
Olsen-P (ppm)	23.2	38.6	5.2
X-K (ppm)	286	337	104
Sand (%)	16	13	61
Silt (%)	39	35	23
Clay (%)	45	52	16
Water Depth (cm)	4.2	1.8	0.4

soils (6.75 vs. 4.20, $P = 0.003$, Table 2). Although not quite statistically significant, there was a suggestion of an ecotype (seed source) effect for seed set, the most fundamental fitness trait (Table 2, $P = 0.08$, Fig. 1).

There was strong evidence of ecotypic differentiation between the alkali and non-alkali populations of *Lolium* (Table 3), in the form of (1) significant seed source effects (for plant height), and (2) significant seed source by soil source interactions (for number of leaves and seed set). Plants grown from alkali seed sources were nearly twice as tall as those grown from the non-alkali seed source, across all three soil sources (25.7 vs. 13.2 cm, $P < 0.001$, Fig. 2a). *Lolium multiflorum*

Table 2 Summary of ANOVA analysis for the effects of seed source and soil source on plant traits of *Hemizonia pungens* and *Lolium multiflorum*, in alkali sink and sink matrix populations (excluding the non-alkali population)

	Seed source		Soil source		Seed * soil source	
	F	P	F	P	F	P
<i>L. multiflorum</i>						
Height	0.43	0.52	2.14	0.15	0.12	0.73
Number of leaves	0.32	0.58	10.39	0.003	0.10	0.75
ln (seed set)	3.20	0.08	0.90	0.35	1.11	0.30
<i>H. pungens</i>						
Height	0.18	0.68	6.99	0.01	0.08	0.78
Number of leaves	0.64	0.43	0.31	0.58	0.31	0.58

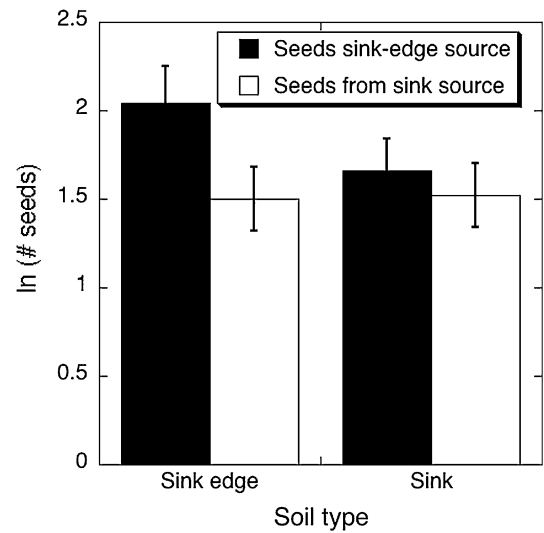
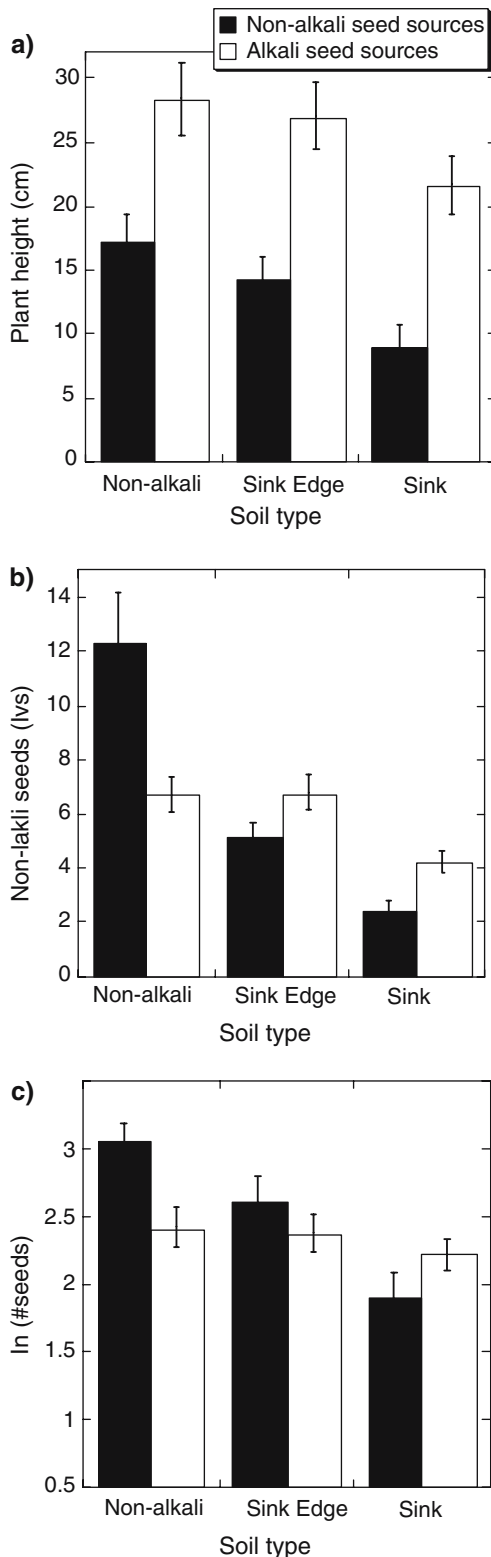


Fig. 1 Seed set (ln-transformed) in *L. multiflorum* grown from seed collected from either sink or sink matrix populations, and grown in the greenhouse in soil collected from either sink or sink matrix sites. Bars represent means \pm one standard error

rum plants grown from the alkali sink seed source were significantly more tolerant of sink and sink matrix soils than were plants grown from the non-alkali seed source: for both number of leaves and seed set (Fig. 2 b,c), there were significant soil source by seed source interaction terms. In both cases, the plants that performed best in either

Table 3 Results of ANOVA analysis comparing the effects of alkali (sink and matrix populations combined) and non-alkali seed sources, and three soil sources (alkali sink, sink matrix, and non-alkali), on plant traits of *L. multiflorum*

Source	df	Sum of squares	F ratio	P
<i>(a) Plant height</i>				
Seed source	1	2777	26.56	0.022
Soil source	2	833	3.98	< 0.0001
Seed*Soil	2	15	0.07	0.93
Error	79	8258		
<i>(b) Number of leaves</i>				
Seed source	1	9.6	1.07	0.30
Soil source	2	510.7	28.51	< 0.0001
Seed*Soil	2	228.0	12.73	< 0.0001
Error	80	1327.0		
<i>(c) ln (seed set)</i>				
Seed source	1	0.67	2.10	0.15
Soil source	2	6.20	9.80	< 0.001
Seed*Soil	2	2.98	4.70	0.01
Error	79	25.00		



◀ **Fig. 2** Plant height (a), number of levels (b), and seed set (c), in *L. multiflorum* grown from seed collected from either alkali (sink and sink matrix) or non-alkali populations, and grown in either sink, sink edge (“sink matrix”), or non-alkali soils. Bars represent means \pm one standard error

alkali sink or non-alkali soils were plants grown from alkali sink and non-alkali seed sources, respectively.

Across all soil and seed sources, *Lolium* seed set was strongly positively correlated with the number of leaves ($r^2 = 0.39, df = 85, P < 0.001$). Seed set was positively correlated with plant height for plants grown from non-alkali seed sources ($r^2 = 0.35, df = 27, P = 0.001$), but negatively correlated with plant height for plants grown from alkali seed sources ($r^2 = 0.19, df = 58, P = 0.001$).

Discussion

The soils of the alkali sink and sink matrix habitats are at or beyond the alkaline limits reported for *L. multiflorum* (7.5–7.8), and are extreme in other soils traits as well (Table 1). Indeed, all seed sources of both the exotic *L. multiflorum* and native *H. pungens* exhibit lower values of fitness traits (plant size and seed set) on the alkali soils than on the non-alkali soils. There was a curious pattern for plant height in *L. multiflorum*, which was consistently greater for the alkali genotypes across all soil types. We suspect that this is an adaptation for the extreme inundation that characterized the alkali sites.

Ideally, all fitness measures would be positively correlated, and vegetative measures would be appropriate indirect measures of more direct fitness measures, such as seed set. However, *Lolium* height was positively correlated with seed set in the non-alkali seed sources, but negatively correlated in the alkali seed sources. One possible explanation for this difference is that in the non-alkali sites, plant height was an indicator of a favourable microsite, and therefore was positively correlated with seed set, but in the alkali sites, ecotypic variation in the resources put into plant height (to escape inundation) comes at the expense of seed set. In any case, this example shows

that different fitness measures can become uncoupled in unexpected ways.

Nonetheless, there was clear evidence of ecotypic differentiation between the alkali and non-alkali populations of *L. multiflorum*. In particular, plants grown from the alkali seed sources performed better on alkali sink soils than did plants grown from the non-alkali seed sources, fulfilling the critical “local vs. foreign” criterion of Kawecki and Ebert (2004). There was even a suggestion of more local alkali ecotypes, because plants grown from seeds from the sink microsites maintained their fecundity on the severe sink soils, whereas plants grown from seeds from the sink matrix microsites experienced a 37% decline in seed set (from 9.0 to 5.7 seeds/plant; Fig. 1). This ecotypic difference was not statistically significant, but was in the direction expected. If confirmed, this would represent ecotypic differentiation on the scale of only a few meters. Although such fine-scale ecotypic variation has been found in similar pools in an insect-pollinated, largely selfing native species (Linhart 1988), for a wind-pollinated, outcrossing species like *L. multiflorum*, one might expect limited ability to maintain ecotypes over very short distances.

We do not know if these forms of ecotypic differentiation are due to recent evolution *in situ*, or to the differential arrival and survival of different genotypes (c.f. Vasquez et al. 2005). The fact that *Lolium multiflorum* has been introduced many times, from a variety of different ecotypes, certainly raises the latter possibility. However, the existence of ecotypes in this study that tolerate pH levels (8.4) far higher than even the extremes reported for *L. multiflorum* in the literature (Hannaway et al. 1999) does suggest local evolutionary change after introduction.

Conservationists and restorationists have long recognized, and taken encouragement from, the fact that edaphically severe sites are often resistant to invasion, and become refuges for native endemic plant species. Recent research, however, has documented the ability of exotic species to invade such sites, albeit slowly (Williamson and Harrison 2002; Gerhardt and Collinge 2003; Zedler and Black 2004). Our study suggests that such invasion may be associated with the arrival

or evolution of specialized ecotypes. This may be ominous news for those trying to conserve, manage, and restore native species in highly invaded ecosystems, and may provide greater impetus to catch invaders early, not merely as a more cost-effective control measure (Dulloo et al. 2002; Chornesky and Randall, 2003), but to slow or even prevent the evolution of local ecotypes.

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References

- Agriculture and Agri-food Canada (1999) Strategies for avoiding herbicide resistance in ryegrasses. Research Letter No. 99–01. Retrieved from http://www.res2.agr.ca/saskatoon/result/letter9901-letter9901_e.htm on 4 December 2005
- Bennington CC, McGraw JB (1995) Natural selection and ecotypic differentiation in *Impatiens pallida*. *Ecological Monographs* 65:303–323
- Berglund ABN, Dahlgren S, Westerbergh A (2004) Evidence for parallel evolution and site-specific selection of serpentine tolerance in *Cerastium alpinum* during the colonization of Scandinavia. *New Phytologist* 161:199–209
- Bradshaw AD (1984) Ecological significances of genetic variation between populations. In: Dirzo R, Sarukhán J (eds) *Perspectives on plant population ecology*. Sinauer Associates Inc., Sunderland, pp 213–228
- Brady KU, Kruckeberg AR, Bradshaw H (2005) Evolutionary ecology of plant adaptation to serpentine soils. *Annual Review of Ecology, Evolution, and Systematics* 36:243–266
- Briggs D, Walters SM (1984) *Plant Variation and Evolution*, 2nd edn. Cambridge University Press, Cambridge
- Brown CS, Rice KJ (2000) The mark of Zorro: effects of the exotic annual grass *Vulpia myuros* on California native perennial grasses. *Restoration Ecology* 8:10–17
- California Exotic Pest Plant Council (1999) Exotic plant pest list. Retrieved from <http://www.ucce.ucdavis.edu/files/filelibrary/5319/4898.pdf> on October 19, 1999
- Cheplick GP (1988) Influence of environment and population origin on survivorship and reproduction in reciprocal transplants of Amphicarpic Peanutgrass (*Amphicarpum purshii*). *American Journal of Botany* 75:1048–1056
- Chornesky EA, Randall JM (2003) The threat of invasive alien species to biological diversity: setting a future course. *Annals of the Missouri Botanical Garden* 90:67–76

- Clausen J, Keck DD, Heisey WM (1940) Experimental studies on the nature of species. I. The effect of varied environments on Western North American plants. Carnegie Institution of Washington Publication No. 520, Washington D.C
- Cox GW (2004) Alien species and evolution. Island Press, Washington DC, USA
- Dulloo ME, Kell SR, Jones CG (2002) Impact and control of invasive alien species on small islands. *International Forestry Review* 4:277–285
- Dyer AR, Rice KJ (1997) Intraspecific and diffuse competition: the response of *Nassella pulchra* in a California grassland. *Ecological Applications* 7:484–492
- Dyer AR, Rice KJ (1999) Effects of competition on resource availability and growth of a California bunchgrass. *Ecology* 80:2697–2710
- Galloway LF, Fenster CB (2000) Population differentiation in an annual legume: local adaptation. *Evolution* 54:1173–1181
- Gelbard JL, Harrison S (2003) Roadless habitats as refuges for native grasslands: interactions with soil, aspect, and grazing. *Ecological Applications* 13:404–415
- Gerhardt F, Collinge SK (2003) Exotic plant invasions of vernal pools in the Central Valley of California, USA. *Journal of Biogeography* 30:1043–1052
- Gram WK, Borer ET, Cottingham KL, Seabloom EW, Boucher VL, Goldwasser L, Micheli F, Kendall BE, Burton RS (2004) Distribution of plants in a California serpentine grassland: are rocky hummocks spatial refuges for native species? *Plant Ecology* 172:159–171
- Hannaway D, Franssen S, Cropper J, Teel M, Chaney M, Griggs T, Halse R, Hart J, Cheeke P, Hansen D, Klinger R, Lane W (1999) Annual ryegrass (*Lolium multiflorum* Per.). Retrieved from <http://www.eesc.orst.edu/agcomwebfile/edmat/html/pnw/pnw501/intro.html> on 5 December 2005
- Harrison S (1999) Native and alien species diversity at the local and regional scales in a grazed California grassland. *Oecologia* 121:99–106
- Heady HF (1988) Valley grassland. In: Barbour MG, Major J (eds) *Terrestrial vegetation of California*. J. Wiley-Interscience, New York, pp 491–514
- Hellmers H, Ashby WC (1958) Growth of native and exotic plants under controlled temperatures and in the San Gabriel Mountains California. *Ecology* 39:416–428
- Hickman JC (ed) (1993) *The Jepson Manual of Higher Plants of California*. University of California Press, Berkeley, CA, USA, 1400 pp
- Joshi J, Schmid B, Caldeira MC, Dimitrakopoulos PG, Good J, Harris R, Hector A, Huss-Danell K, Jumpponen A, Minns A, Mulder CPH, Pereira JS, Prinz A, Sherer-Lorenzen M, Siamantziouras ASD, Terry AC, Troumbis AY, Lawton JH (2001) Local adaptation enhances performance of common plant species. *Ecology Letters* 4:536–544
- Kawecki T, Ebert D (2004) Conceptual issues in local adaptation. *Ecology Letters* 7:1225–1241
- Kay BL, Love RM, Slayback RD (1981) Revegetation with native grasses: a disappointing history. *Fremontia* 9:11–15
- Kruckeberg AR (1951) Intraspecific variability in the response of certain native plant species to serpentine soil. *American Journal of Botany* 38:408–419
- Leiss KA, Müller-Schärer H (2001) Performance of reciprocally sown populations of *Senecio vulgaris* from ruderal and agricultural habitats. *Oecologia* 128:210–216
- Linhart YB (1988) Intrapopulation differentiation in annual plants 3. The contrasting effects of intraspecific and interspecific competition. *Evolution* 42:1047–1064
- Marty JT (2005) Effects of cattle grazing on diversity in ephemeral wetlands. *Conservation Biology* 19:1626–1632
- McGraw JB, Chapin FS III (1989) Competitive ability and adaptation to fertile and infertile soils in two *Eriophorum* species. *Ecology* 70:736–749
- Montalvo AM, Ellstrand NC (2000) Transplantation of the subshrub *Lotus scoparius*: testing the home-site advantage hypothesis. *Conservation Biology* 14:1034–1045
- Prasada R, Power JF (1997) *Soil Fertility Management for Sustainable Agriculture*. Lewis Publishers, NY, NY
- Primack RB, Kang H (1989) Measuring fitness and natural selection in wild plant populations. *Annual Review of Ecology and Systematics* 20:367–396
- Vasquez EA, Glenn EP, Brown JJ, Guntenspergen GR, Nelson SG (2005) Salt tolerance underlies the cryptic invasion of North American salt marshes by an introduced haplotype of the common reed *Phragmites australis* (Poaceae). *Marine Ecology-Progress Series* 298:1–8
- Vitousek PM., D'Antonio CM, Loope LL, Rejmanek M, Westbrooks R (1997) Introduced species: a significant component of human-caused global change. *New Zealand Journal of Ecology* 21:1–16
- Wilcove DS, Rothstein D, Dubrow J, Phillips A, Losos E (1998) Quantifying threats to imperiled species in the United States. *BioScience* 48:607–615
- Williamson J, Harrison S (2002) Biotic and abiotic limits to the spread of exotic revegetation species. *Ecological Applications* 12:40–51
- Zedler PH, Black C (2004) Exotic plant invasions in an endemic-rich habitat: The spread of an introduced Australian grass, *Agrostis avenacea* J. F. Gmel., in California vernal pools. *Austral Ecology* 29:537–546